



# NAD Kinase Microplate Assay Kit User Manual

Catalog # FTA0040

(Version 1.1C)

Detection and Quantification of NAD Kinase (NADK) Activity in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



I. INTRODUCTION	2
II. KIT COMPONENTS	3
III. MATERIALS REQUIRED BUT NOT PROVIDED	3
IV. SAMPLE PREPARATION	4
V. ASSAY PROCEDURE	5
VI. CALCULATION	6
VII. TYPICAL DATA	7
VIII. TECHNICAL SUPPORT	7
IX NOTES	7



## I. INTRODUCTION

NAD kinase catalyzes the transfer of a phosphate group from ATP to NAD<sup>+</sup> to generate NADP<sup>+</sup>, which in its reduced form acts as an electron donor for biosynthetic reactions. NADP<sup>+</sup> is an essential coenzyme in metabolism and provides reducing power to biosynthetic processes such as fatty acid biosynthesis.

The assay is initiated with the enzymatic hydrolysis of the NAD<sup>+</sup> by NADK. The enzyme catalysed reaction products can be measured at a colorimetric readout at 600 nm.



## **II. KIT COMPONENTS**

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Substrate Diluent	16 ml x 1	4 °C
Substrate	Powder x 1	-20 °C
Enzyme	Powder x 1	-20 °C
Dye Reagent	Powder x 1	-20 °C, keep in dark
Stop Solution	20 ml x 1	4 °C
Dissolution Buffer	30 ml x 1	4 °C
Standard	Powder x 1	-20 °C, keep in dark
Technical Manual	1 Manual	

## Note:

Substrate: add 8 ml Substrate Diluent to dissolve before use.

**Enzyme**: add 10 ml distilled water to dissolve before use, mix, store at 4°C.

**Dye Reagent**: add 1 ml distilled water to dissolve before use, mix, store at 4°C.

Standard: add 1 ml distilled water to dissolve, mix; then add 25  $\mu l$  solution into 975

 $\mu$ l distilled water, mix. The concentration will be 50  $\mu$ mol/L.

## III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 570 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer



#### IV. SAMPLE PREPARATION

## 1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10<sup>6</sup> cell or bacteria, sonicate (with power 20%, sonication 3s, intervation 10s, repeat 30 times); centrifuged at 8,000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

# 2. For tissue samples

Weigh 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

# 3. For serum or plasma samples

Detect directly.



## V. ASSAY PROCEDURE

Add following reagents in the microcentrifuge tubes:

Reagent	Sample	Control	Standard	Blank		
Sample	20 μΙ	1	-			
Assay Buffer	-	20 μΙ				
Substrate	80 μΙ	80 μΙ				
Mix, put it in the oven, 37 °C for 10 minutes. Then put it in boiling water for 2						
minutes. When cold, centrifuged at 10000g, room temperature for 10 minutes, take						
the supernatant into the new microcentrifuge tubes.						
The supernatant	20 μΙ	20 μΙ				
Standard			20 μΙ			
Distilled water				20 μΙ		
Enzyme	90 μΙ	90 μΙ	90 μΙ	90 μΙ		
Dye Reagent	10 μΙ	10 μΙ	10 μΙ	10 μΙ		
Mix, keep them in dark for 2 minutes at room temperature.						
Stop Solution	200 μΙ	200 μΙ	200 μΙ	200 μΙ		
Mix, stand at room temperature for 1 minutes, centrifuged at 20,000g for 5						
minutes, discard the supernatant after centrifugation.						
Dissolution Buffer	300 μΙ	300 μΙ	300 μΙ	300 μΙ		
Add 200 µl solution into the microplate, record absorbance measured at 570 nm.						

## Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.



## VI. CALCULATION

**Unit Definition:** One unit of NADK activity is the enzyme that generates 1 nmol of NADP per minute.

1. According to the protein concentration of sample

NADK (U/mg) = 
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) /$$

$$(V_{Sample} \times C_{Protein}) / T \times 10$$

$$= 50 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / C_{Protein}$$

2. According to the weight of sample

NADK (U/g) = 
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) /$$

$$(V_{Sample} \times W / V_{Assay}) / T$$

$$= 50 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / W$$

3. According to the quantity of cells or bacteria

NADK (U/10<sup>4</sup>) = (C<sub>Standard</sub> × V<sub>Standard</sub>) × (OD<sub>Sample</sub> - OD<sub>Control</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / 
$$(V_{Sample} \times N / V_{Assay}) / T$$
=  $50 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / N$ 

C<sub>Protein</sub>: the protein concentration, mg/ml;

 $C_{Standard}$ : the standard concentration, 50  $\mu$ mol/L = 50 nmol/ml

W: the weight of sample, g;

V<sub>Sample</sub>: the volume of sample, 0.02 ml;

V<sub>Assay</sub>: the volume of Assay buffer, 1 ml;

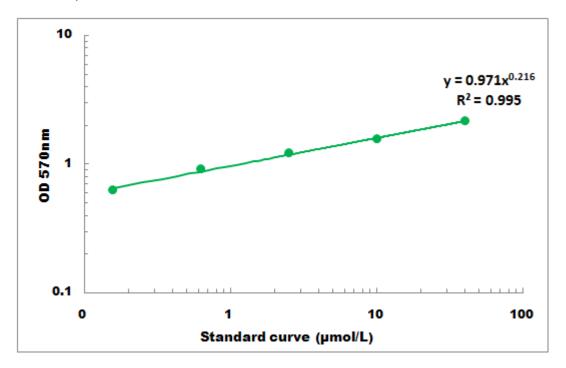
N: the quantity of cell or bacteria,  $N \times 10^4$ ;

T: the reaction time, 10 minutes.



## VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.1 μmol/L - 50 μmol/L

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

## IX. NOTES