



Monodehydroascorbate Reductase Microplate Assay Kit User Manual

Catalog # FTA0052

(Version 1.2C)

Detection and Quantification of Monodehydroascorbate Reductase (MDAR) Activity in Tissue extracts, Cell lysate Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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I. INTRODUCTION

In plants, the monodehydroascorbate reductase (MDAR) is an enzymatic component of the glutathione-ascorbate cycle that is one of the major antioxidant systems of plant cells for the protection against the damages produced by reactive oxygen species (ROS). The MDAR activity has been described in several cell compartments, such as chloroplasts, cytosol, mitochondria, glyoxysomes, and leaf peroxisomes. The assay is initiated with the enzymatic catalysis of the NADH by MDAR. NADH can be measured at a colorimetric readout at 340 nm.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30ml x 4	4 °C
Reaction Buffer	20 ml x 1	4 °C
Enzyme	Powder x 1	-20 °C, keep in dark
Substrate	Powder x 1	-20 °C
Standard	Powder x 1	-20 °C
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Note:

Enzyme: add 1 ml Reaction Buffer to dissolve before use.

Substrate: add 1 ml Reaction Buffer to dissolve before use.

Standard: add 1 ml distilled water to dissolve before use; then add 0.2 ml into 0.8 ml

distilled water, the concentration will be 400 µmol/L.

III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 340 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Ice



IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonication 3s, intervation 10s, repeat 30 times); centrifuged at 10000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 10000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.



V. ASSAY PROCEDURE

Warm all reagents to room temperature before use.

Add following reagents in the microplate:

Reagent	Sample	Standard	Blank		
Reaction Buffer	170 μΙ				
Enzyme	10 μΙ				
Substrate	10 μΙ				
Mix.					
Standard		200 μΙ			
Distilled water			200 μΙ		
Sample	10 μΙ				
Mix. measured at 340 nm and record the absorbance of 10th second and 130th					

Note:

second.

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.



VI. CALCULATION

Unit Definition: One unit of MDAR is defined as the enzyme oxidize 1 nmol NADH per minute.

1. According to the protein concentration of sample

MDAR (U/mg) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} \times C_{Protein}) / T$$

$$= 4000 \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / C_{Protein}$$

2. According to the weight of sample

MDAR (U/g) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank})$$

 $/ (V_{Sample} \times W / V_{Assay}) / T$
= $4000 \times (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / W$

3. According to the quantity of cells or bacteria

MDAR (U/10⁴) = (C_{Standard} × V_{Standard}) × (OD_{Sample(10S)} - OD_{Sample(130S)}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} × N / V_{Assay}) / T
$$= 4000 \times (ODSample(10S) - ODSample(130S)) / (ODStandard - ODBlank) / N$$

4. According to the volume of sample

$$\begin{aligned} \text{MDAR (U/mI)} &= \left(\text{C}_{\text{Standard}} \times \text{V}_{\text{Standard}} \right) \times \left(\text{OD}_{\text{Sample(10S)}} - \text{OD}_{\text{Sample(130S)}} \right) / \left(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}} \right) / \text{V}_{\text{Sample}} / \text{T} \\ &= 4000 \times \left(\text{OD}_{\text{Sample(10S)}} - \text{OD}_{\text{Sample(130S)}} \right) / \left(\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}} \right) \end{aligned}$$

 $C_{Standard}$: the standard concentration, 400 µmol/L = 400 nmol/ml;

 $V_{Standard}$: the volume of standard, 200 μ l = 0.2 ml;

C_{Protein}: the protein concentration, mg/ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria, $N \times 10^4$;

V_{Sample}: the volume of sample, 0.01 ml;

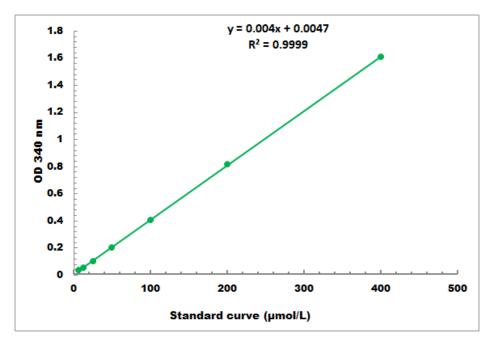
V_{Assay}: the volume of Assay buffer, 1 ml;

T: the reaction time, 2 minutes.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 4 μmol/L - 400 μmol/L

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES