



Basic Xylanase Microplate Assay Kit User Manual

Catalog # FTA0090

(Version 1.2C)

Detection and Quantification of Basic Xylanase (BAX) Activity in Animal feeds, Enzyme preparations, Bread improver mixtures and other materials Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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I. INTRODUCTION

Xylanase (EC 3.2.1.8) is the name given to a class of enzymes which degrade the linear polysaccharide beta-1,4-xylan into xylose, thus breaking down hemicellulose, one of the major components of plant cell walls. As such, it plays a major role in micro-organisms thriving on plant sources for the degradation of plant matter into usable nutrients. Xylanases are produced by fungi, bacteria, yeast, marine algae, protozoans, snails, crustaceans, insect, seeds, etc., (mammals do not produce xylanases).

The assay is initiated with the enzymatic hydrolysis of the xylan by basic xylanase.

The enzyme catalysed reaction products react with 3,5-dinitrosalicylic acid, and can be measured at a colorimetric readout at 540 nm.



II. KIT COMPONENTS

Component	Volume	Storage	
96-Well Microplate	1 plate		
Assay Buffer	30 ml x 4	4 °C	
Substrate	Powder x 1	4 °C	
Dye Reagent	10 ml x 1	4 °C, keep in dark	
Standard (2.5 mmol/L)	1 ml x 1	4 °C	
Plate Adhesive Strips	3 Strips		
Technical Manual	1 Manual		

Note:

Substrate: add 10 ml Assay Buffer to dissolve before use, store at 4 °C.

III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 540 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Convection oven



IV. SAMPLE PREPARATION

For animal feeds, enzyme preparations, bread improver mixtures samples
 Weigh out 0.1 g sample, homogenize with 1 ml Assay buffer, centrifuged at 8,000g
 C for 20 minutes, take the supernatant into a new centrifuge tube for detection.

2. For liquid sample

Add 0.1 ml sample into 0.9 ml Assay buffer, centrifuged at 8,000g 4 °C for 20 minutes, take the supernatant into a new centrifuge tube for detection.



V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Control	Standard	Blank		
Sample	10 μΙ					
Distilled water		10 μΙ				
Substrate	90 μΙ	90 μΙ				
Mix, put it in the oven, 50 °C for 10 minutes.						
Standard			100 μΙ			
Distilled water				100 μΙ		
Dye Reagent	100 μΙ	100 μΙ	100 μΙ	100 μΙ		

Mix, put it into the convection oven, 90 °C for 10 minutes, when cold record absorbance measured at 540 nm.

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.



VI. CALCULATION

Unit Definition: One unit of Basic Xylanase activity is defined as the enzyme generates 1 μ mol of reducing sugar per hour at 50 °C, pH9.0.

1. According to the weight of sample

BAX (U/g) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} \times W / V_{Assay}) / T$$

$$= 2.5 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / W$$

2. According to the volume of sample

BAX (U/mI) =
$$(C_{Standard} \times V_{Standard}) \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) /$$

$$(V_{Sample} \times V / V_{Assay}) / T$$

$$= 2.5 \times (OD_{Sample} - OD_{Control}) / (OD_{Standard} - OD_{Blank}) / V$$

 $C_{Standard}$: the concentration of standard, 2.5 mmol/L = 2.5 μ mol/ml;

W: the weight of sample, g;

V: the volume of sample, ml;

V_{Standard}: the volume of standard, 0.1 ml;

V_{Sample}: the volume of sample, 0.01 ml;

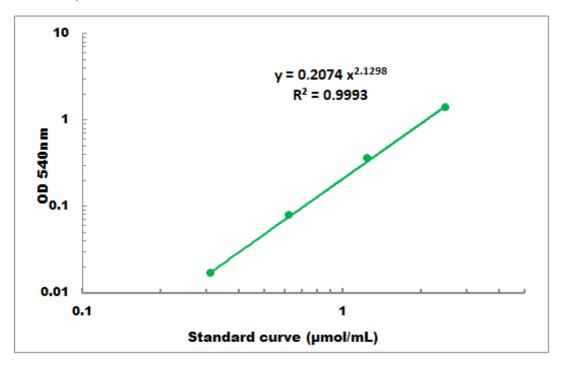
V_{Assay}: the volume of Assay buffer in sample preparation, 1 ml;

T: the reaction time, 10 minutes.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.25 mmol/L - 2.5 mmol/L

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES