



# Glycogen Microplate Assay Kit User Manual

Catalog # FTA0128

(Version 1.2A)

Detection and Quantification of Glycogen Content in Tissue extracts, Cell lysate, Cell culture media and Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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## I. INTRODUCTION

Glycogen is a multibranched polysaccharide of glucose that serves as a form of energy storage in humans, animals, and fungi. The polysaccharide structure represents the main storage form of glucose in the body.

In humans, glycogen is made and stored primarily in the cells of the liver and the muscles, hydrated with three or four parts of water. Glycogen functions as the secondary long-term energy storage, with the primary energy stores being fats held in adipose tissue. Muscle glycogen is converted into glucose by muscle cells, and liver glycogen converts to glucose for use throughout the body including the central nervous system.

Glycogen Microplate Assay Kit is a sensitive assay for determining Glycogen in various samples. Glycogen concentration is determined by anthrone. The reaction products can be measured at a colorimetric readout at 620 nm.



## **II. KIT COMPONENTS**

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	30 ml x 4	4 °C
Dye Reagent	Powder x 1	4 °C
Dye Reagent Diluent	15 ml x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

## Note:

**Standard**: add 1 ml distilled water to dissolve before use, then add 20  $\mu$ l into 980  $\mu$ l distilled water, the concentration will be 200  $\mu$ g/ml.

Dye Reagent: add 15 ml Dye Reagent Diluent to dissolve before use, store at 4 °C.

**Dye Reagent Diluent**: be careful, it is strong acid.

# III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 620 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. Ice



#### IV. SAMPLE PREPARATION

# 1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonication 3s, intervation 10s, repeat 30 times); centrifuged at 10,000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

# 2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 10,000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

# 3. For liquid samples

Add 0.9 ml Assay buffer into 0.1 ml liquid sample, centrifuged at 10,000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.



## V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank
Sample	50 μΙ		
Standard		50 μΙ	
Distilled water			50 μΙ
Dye Reagent	150 μΙ	150 μΙ	150 μΙ

Mix, put the microplate into the oven, incubate at 90 °C for 15 minutes, when cold record absorbance measured at 620 nm.

#### Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.



## VI. CALCULATION

1. According to the protein concentration of sample

Glycogen (
$$\mu$$
g/mg) = 1.11 × ( $C_{Standard}$  ×  $V_{Standard}$ ) × ( $OD_{Sample}$  -  $OD_{Blank}$ ) / ( $OD_{Standard}$  -  $OD_{Blank}$ ) / ( $V_{Sample}$  ×  $C_{Protein}$ )
$$= 222 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / C_{Protein}$$

2. According to the weight of sample

Glycogen (
$$\mu$$
g/g) = 1.11 × ( $C_{Standard}$  ×  $V_{Standard}$ ) × ( $OD_{Sample}$  -  $OD_{Blank}$ ) / ( $OD_{Standard}$  -  $OD_{Blank}$ ) / ( $V_{Sample}$  × W/  $V_{Assay}$ )
$$= 222 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / W$$

3. According to the quantity of cells or bacteria

Glycogen (
$$\mu$$
g/10<sup>4</sup>) = 1.11 × ( $C_{Standard}$  ×  $V_{Standard}$ ) × ( $OD_{Sample}$  -  $OD_{Blank}$ ) / ( $OD_{Standard}$  -  $OD_{Blank}$ ) / ( $V_{Sample}$  × N/  $V_{Assay}$ ) = 222 × ( $OD_{Sample}$  -  $OD_{Blank}$ ) / ( $OD_{Standard}$  -  $OD_{Blank}$ ) / N

4. According to the volume of sample

Glycogen (
$$\mu$$
g/ml) = 1.11 × (C<sub>Standard</sub> × V<sub>Standard</sub>) × (OD<sub>Sample</sub> - OD<sub>Blank</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>) / V<sub>Sample</sub> = 222 × (OD<sub>Sample</sub> - OD<sub>Blank</sub>) / (OD<sub>Standard</sub> - OD<sub>Blank</sub>)

1.11: the constant for converting glucose content into glycogen content;

C<sub>Protein</sub>: the protein concentration, mg/ml;

C<sub>Standard</sub>: the protein concentration, 200 µg/ml;

W: the weight of sample, g;

N: the quantity of cell or bacteria,  $N \times 10^4$ ;

 $V_{Standard}$ : the volume of standard, 50 µl;

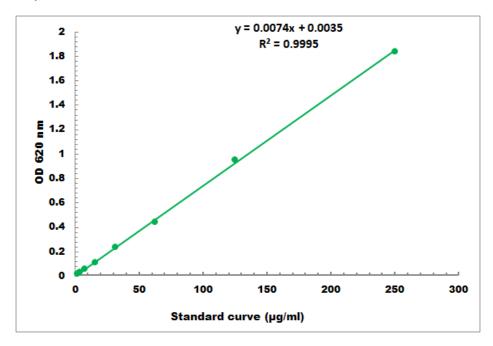
 $V_{Sample}$ : the volume of sample, 50  $\mu$ l.

V<sub>Assay</sub>: the volume of Assay buffer, 1 ml.



# VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 2 μg/ml - 200 μg/ml

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

# IX. NOTES