



TBARS Microplate Assay Kit User Manual

Catalog # FTA0145

(Version 1.2A)

Detection and Quantification of Thiobarbituric Acid Reactive Substances (TBARS) Content in Urine, Serum, Plasma, Tissue extracts, Cell lysate, Cell culture media, Other biological fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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I. INTRODUCTION

Oxidative attack of essential cell components by reactive oxygen species has been associated with several human diseases, such as atherosclerosis, cardiovascular diseases, diabetes, liver disorders, and inflammatory rheumatic diseases.

Thiobarbituric Acid Reactive Substances (TBARS) are low-molecular-weight end products (mainly malondialdehyde, MDA) that are formed during the decomposition of lipid peroxidation products. Increased levels of TBARS have been demonstrated in these diseases

TBARS Microplate Assay Kit is a sensitive assay for determining TBARS concentration in various samples. TBARS concentration is based on the reaction of TBARS with thiobarbituric acid (TBA) to form a pink colored product. The color intensity at 535 nm is directly proportional to TBARS concentration in the sample.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer	10 ml x 1	RT
Dye Reagent	15 ml x 1	RT
Standard (25 μmol/L)	1 ml x 1	RT
Technical Manual	1 Manual	

III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 535 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Centrifuge
- 7. Timer
- 8. PBS



IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml PBS for 5×10^6 cell or bacteria, sonicate (with power 20%, sonication 3s, intervation 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes. Transfer 100 μ l sample into a micro-centrifuge tube. Then add 100 μ l Assay buffer into a micro-centrifuge tube. Incubate for 5 minutes on ice; centrifuged at 12000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml PBS on ice, centrifuged at 8000g 4 °C for 10 minutes. Transfer 100 μ l sample into a micro-centrifuge tube. Then add 100 μ l Assay buffer into a micro-centrifuge tube. Incubate for 5 minutes on ice; centrifuged at 12000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For serum, plasma or other biological fluids samples

Transfer 100 μ l sample into a micro-centrifuge tube. Then add 100 μ l Assay buffer into a micro-centrifuge tube. Incubate for 5 minutes on ice; centrifuged at 12000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.



V. ASSAY PROCEDURE

Warm the Dye Reagent to 37 °C before use.

Add following reagents in the microcentrifuge tube:

Reagent	Sample	Standard	Blank		
Sample	150 μΙ				
Standard		150 μΙ			
Distilled water			150 μΙ		
Dye Reagent	150 μΙ	150 μΙ	150 μΙ		
Shake and mix, put them into boiling water bath for 10 minutes. When cold, add the					
Supernatant into the microplate.					
Supernatant	200 μΙ	200 μΙ	200 μΙ		
Record absorbance measured at 535 nm.					

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.



VI. CALCULATION

1. According to the protein concentration of sample

TBARS (
$$\mu$$
mol/mg) = C_{Standard} × V_{Standard} × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} × C_{Protein}) × 2
$$= 0.05 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / C_{Protein}$$

2. According to the weight of sample

TBARS (
$$\mu$$
mol/g) = C_{Standard} × V_{Standard} × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} × W / V_{PBS}) × 2
$$= 0.05 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / W$$

3. According to the volume of sample

TBARS (
$$\mu$$
mol/ml) = C_{Standard} × V_{Standard} × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / V_{Sample} × 2
$$= 0.05 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank})$$

4. According to the quantity of cell or bacteria

TBARS (
$$\mu$$
mol/10⁴ cell) = C_{Standard} × V_{Standard} × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank})
/ (V_{Sample} × N / V_{PBS}) × 2
= 0.05 × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / N

 $C_{Standard}$: the standard concentration, 25 µmol/L = 0.025 µmol/ml;

V_{Standard}: the volume of standard, 0.15 ml;

C_{Protein}: the protein concentration, mg/ml;

W: the weight of sample, g;

V_{Sample}: the volume of sample, 0.15 ml;

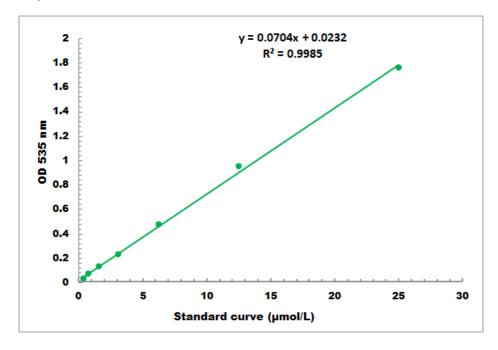
V_{PBS}: the volume of PBS, 1 ml;

N: the quantity of cell or bacteria, $N \times 10^4$.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.5 μmol/L - 25 μmol/L

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES