



Lysine Microplate Assay Kit User Manual

Catalog # FTA0184

(Version 1.2A)

Detection and Quantification of Lysine Content in Serum, Plasma,
Tissue extracts, Cell lysate, Cell culture media and Other biological
fluids Samples.

For research use only. Not for diagnostic or therapeutic procedures.



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I. INTRODUCTION

Lysine (symbol Lys or K) is an α -amino acid that is used in the biosynthesis of proteins. It contains an α -amino group (which is in the protonated –NH3+ form under biological conditions), an α -carboxylic acid group (which is in the deprotonated –COO– form under biological conditions), and a side chain lysyl ((CH2)4NH2), classifying it as a basic, charged (at physiological pH), aliphatic amino acid. The human body cannot synthesize lysine, so it is essential in humans and must be obtained from the diet. In organisms that synthesise lysine, it has two main biosynthetic pathways, the diaminopimelate and α -aminoadipate pathways, which employ different enzymes and substrates and are found in different organisms. Lysine catabolism occurs through one of several pathways, the most common of which is the saccharopine pathway.

Lysine plays several roles in humans, most importantly proteinogenesis, but also in the crosslinking of collagen polypeptides, uptake of essential mineral nutrients, and in the production of carnitine, which is key in fatty acid metabolism. Lysine is also often involved in histone modifications, and thus, impacts the epigenome. The ϵ -amino group often participates in hydrogen bonding and as a general base in catalysis. The ϵ -ammonium group (NH3+) is attached to the fourth carbon from the α -carbon, which is attached to the carboxyl (C=OOH) group.

Lysine react with ninhydrin, the reaction products can be measured at a colorimetric readout at 478 nm.



II. KIT COMPONENTS

Component	Volume	Storage
96-Well Microplate	1 plate	
Assay Buffer I	30 ml x 2	4 °C
Assay Buffer II	30 ml x 2	4 °C
Reaction Buffer	5 ml x 1	4 °C
Inhibitor	2 ml x 1	4 °C
Dye Reagent	Powder x 1	4 °C
Standard	Powder x 1	4 °C
Plate Adhesive Strips	3 Strips	
Technical Manual	1 Manual	

Note:

Dye Reagent: add 5 ml distilled water to dissolve before use, store at 4 °C.

Standard: add 1 ml distilled water to dissolve before use, then then add 125 µl into

875 μ l distilled water, mix; the concentration will be 5 mmol/L.

III. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Microplate reader to read absorbance at 478 nm
- 2. Distilled water
- 3. Pipettor, multi-channel pipettor
- 4. Pipette tips
- 5. Mortar
- 6. Convection oven
- 7. Centrifuge
- 8. Timer



IV. SAMPLE PREPARATION

1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for 5×10^6 cell or bacteria, sonicate (with power 20%, sonication 3s, intervation 10s, repeat 30 times); centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

2. For tissue samples

Weigh 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

3. For serum or plasma samples

Add 1 ml Assay buffer for 0.1 ml serum or plasma; mix; centrifuged at 8000g 4 °C for 10 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.



V. ASSAY PROCEDURE

Add following reagents into the microplate:

Reagent	Sample	Standard	Blank
Sample	80 μΙ		
Standard		80 μΙ	
Distilled water			80 μΙ
Reaction Buffer	50 μΙ	50 μΙ	50 μΙ
Inhibitor	20 μΙ	20 μΙ	20 μΙ
Dye Reagent	50 μΙ	50 μΙ	50 μΙ

Mix, put it into the convection oven, 90 °C for 20 minutes, record absorbance measured at 478 nm.

Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) The concentrations can vary over a wide range depending on the different samples. For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range.



VI. CALCULATION

1. According to the protein concentration of sample

Lysine (
$$\mu$$
mol/ml) = (C_{Standard} × V_{Standard}) × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} × C_{Protein})
$$= 5 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / C_{Protein}$$

2. According to the quantity of cells or bacteria

Lysine (
$$\mu$$
mol/10⁴ cell) = (C_{Standard} × V_{Standard}) × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / (V_{Sample} × N/ V_{Assay})
$$= 5 \times (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / N$$

3. According to the weight of sample

Lysine (
$$\mu$$
mol/g) = (C_{Standard} × V_{Standard}) × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank})/

(V_{Sample} × W/ V_{Assay})

= 5 × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / W

4. According to the volume of sample

Lysine (
$$\mu$$
mol/ml) = (C_{Standard} × V_{Standard}) × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank}) / V_{Sample} = 5 × (OD_{Sample} - OD_{Blank}) / (OD_{Standard} - OD_{Blank})

 $C_{Standard}$: the standard concentration, 5 mmol/L = 5 μ mol/ml;

V_{Sample}: the volume of the sample, 0.08 ml;

V_{Standard}: the volume of the standard, 0.08 ml;

V_{Assay}: the volume of Assay buffer, 1 ml;

C_{Protein}: the protein concentration, µmol/ml;

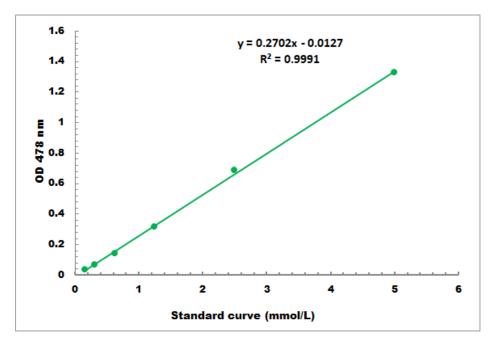
W: the weight of sample, g;

N: the quantity of cell or bacteria, $N \times 10^4$.



VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.1 mmol/L - 5 mmol/L

VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to www.cohesionbio.com or contact us at techsupport@cohesionbio.com

IX. NOTES