



# **Nitrite Reductase Microplate Assay Kit User Manual**

**Catalog # FTA0198**

(Version 1.2A)

Detection and Quantification of Nitrite Reductase (NiR) Activity in  
Tissue extracts, Cell lysate, Cell culture media and Other biological  
fluids Samples.

**For research use only. Not for diagnostic or therapeutic procedures.**

|   |   |
|---|---|
| I. INTRODUCTION.....                          | 2 |
| II. KIT COMPONENTS.....                       | 3 |
| III. MATERIALS REQUIRED BUT NOT PROVIDED..... | 3 |
| IV. SAMPLE PREPARATION.....                   | 4 |
| V. ASSAY PROCEDURE.....                       | 5 |
| VI. CALCULATION.....                          | 6 |
| VII. TYPICAL DATA.....                        | 7 |
| VIII. TECHNICAL SUPPORT.....                  | 7 |
| IX. NOTES.....                                | 7 |

## I. INTRODUCTION

Nitrite reductase refers to any of several classes of enzymes that catalyze the reduction of nitrite. There are two classes of NIR's. A multi haem enzyme reduces  $\text{NO}_2^-$  to a variety of products. Copper containing enzymes carry out a single electron transfer to produce nitric oxide.

Nitrite Reductase Microplate Assay Kit is a sensitive assay for determining Nitrite Reductase activity in various samples. Nitrite reductase can reduce  $\text{NO}_2^-$  to NO.  $\text{NO}_2^-$  can react with dye reagent, and can be measured at a colorimetric readout at 540 nm. The reduction of  $\text{NO}_2^-$  is proportional to the nitrite reductase activity.

## II. KIT COMPONENTS

| Component           | Volume     | Storage |
|---------------------|------------|---------|
| 96-Well Microplate  | 1 plate    |         |
| Assay Buffer        | 30 ml x 4  | 4 °C    |
| Reaction Buffer     | 6 ml x 1   | 4 °C    |
| Substrate           | Powder x 1 | 4 °C    |
| Stop Solution       | 5 ml x 1   | 4 °C    |
| Dye Reagent         | Powder x 1 | 4 °C    |
| Dye Reagent Diluent | 10 ml x 1  | 4 °C    |
| Standard            | Powder x 1 | 4 °C    |
| Technical Manual    | 1 Manual   |         |

### Note:

**Substrate:** add 2 ml distilled water to dissolve before use.

**Dye Reagent:** add 10 ml Dye Reagent Diluent to dissolve before use.

**Standard:** add 1 ml distilled water to dissolve before use; then add 20 µl into 980 µl distilled water. The concentration will be 2 mmol/L.

## III. MATERIALS REQUIRED BUT NOT PROVIDED

1. Microplate reader to read absorbance at 540 nm
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Ice
7. Centrifuge
8. Timer

#### IV. SAMPLE PREPARATION

##### 1. For cell and bacteria samples

Collect cell or bacteria into centrifuge tube, discard the supernatant after centrifugation, add 1 ml Assay buffer for  $5 \times 10^6$  cell or bacteria, sonicate (with power 20%, sonication 3s, interval 10s, repeat 30 times); centrifuged at 4000g 4 °C for 5 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

##### 2. For tissue samples

Weigh out 0.1 g tissue, homogenize with 1 ml Assay buffer on ice, centrifuged at 4000g 4 °C for 5 minutes, take the supernatant into a new centrifuge tube and keep it on ice for detection.

##### 3. For liquid samples

Detect directly.

## V. ASSAY PROCEDURE

Add following reagents into the microplate:

| Reagent   | Sample      | Control     | Standard    | Blank       |
|---|-------------|-------------|-------------|-------------|
| Reaction Buffer   | 60 $\mu$ l  | 60 $\mu$ l  | 60 $\mu$ l  | 60 $\mu$ l  |
| Substrate   | 20 $\mu$ l  | 20 $\mu$ l  | --          | --          |
| Sample  | 20 $\mu$ l  | --          | --          | --          |
| Standard  | --          | --          | 20 $\mu$ l  | --          |
| Distilled water   | --          | 20 $\mu$ l  | 20 $\mu$ l  | 40 $\mu$ l  |
| Mix, put it in the oven, 37 °C for 30 minutes.  |             |             |             |             |
| Stop Solution   | 50 $\mu$ l  | 50 $\mu$ l  | 50 $\mu$ l  | 50 $\mu$ l  |
| Mix, centrifuged at 10,000g 4 °C for 10 minutes, add the supernatant into the microplate. |             |             |             |             |
| Supernatant   | 100 $\mu$ l | 100 $\mu$ l | 100 $\mu$ l | 100 $\mu$ l |
| Dye Reagent   | 100 $\mu$ l | 100 $\mu$ l | 100 $\mu$ l | 100 $\mu$ l |
| Mix, record absorbance measured at 540 nm.  |             |             |             |             |

### Note:

- 1) Perform 2-fold serial dilutions of the top standards to make the standard curve.
- 2) For unknown samples, we recommend doing a pilot experiment & testing several doses to ensure the readings are within the standard curve range. If the enzyme activity is lower, please add more sample into the reaction system; or increase the reaction time; if the enzyme activity is higher, please dilute the sample, or decrease the reaction time.

## VI. CALCULATION

**Unit Definition:** One unit of NiR activity is defined as the enzyme reduce 1  $\mu\text{mol}$  of  $\text{NO}_2^-$  per hour.

### 1. According to the protein concentration of sample

$$\begin{aligned}\text{NiR (U/mg)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / \\ &\quad (C_{\text{Protein}} \times V_{\text{Sample}}) / T \\ &= 4 \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / C_{\text{Protein}}\end{aligned}$$

### 2. According to the weight of sample

$$\begin{aligned}\text{NiR (U/g)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / (V_{\text{Sample}} \\ &\quad \times W / V_{\text{Assay}}) / T \\ &= 4 \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / W\end{aligned}$$

### 3. According to the quantity of cells or bacteria

$$\begin{aligned}\text{NAG (U/10}^4\text{)} &= (C_{\text{Standard}} \times V_{\text{Standard}}) \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / \\ &\quad (V_{\text{Sample}} \times N / V_{\text{Assay}}) / T \\ &= 4 \times (\text{OD}_{\text{Control}} - \text{OD}_{\text{Sample}}) / (\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}) / N\end{aligned}$$

$C_{\text{Protein}}$ : the protein concentration, mg/ml;

$C_{\text{Standard}}$ : the concentration of standard, 2 mmol/L = 2  $\mu\text{mol/ml}$ ;

W: the weight of sample, g;

N: the quantity of cell or bacteria,  $N \times 10^4$ ;

$V_{\text{Standard}}$ : the volume of standard, 0.02 ml;

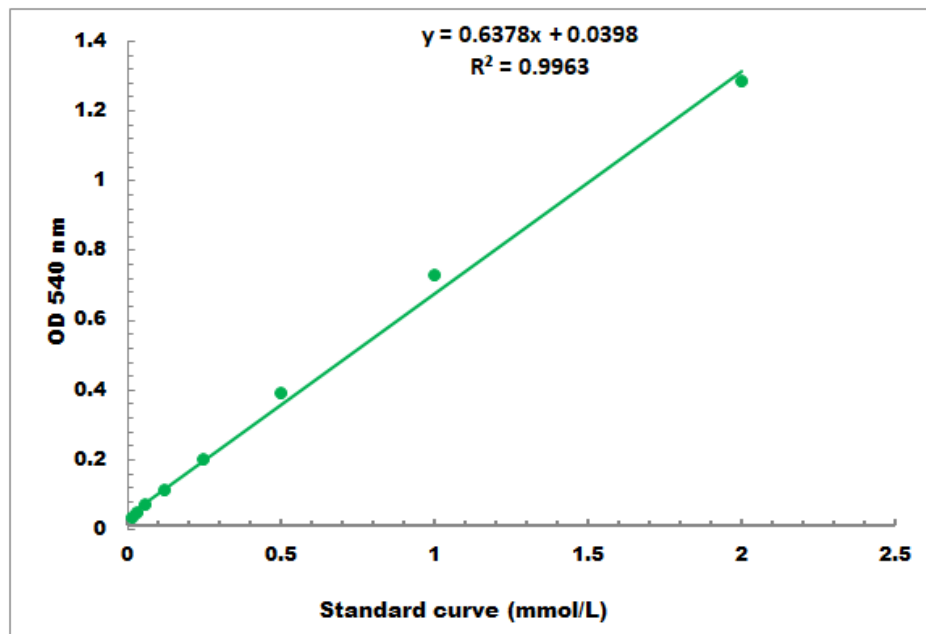
$V_{\text{Sample}}$ : the volume of sample, 0.02 ml;

$V_{\text{Assay}}$ : the volume of Assay buffer, 1 ml;

T: the reaction time, 30 minutes = 0.5 hour.

## VII. TYPICAL DATA

The standard curve is for demonstration only. A standard curve must be run with each assay.



Detection Range: 0.02 mmol/L - 2 mmol/L

## VIII. TECHNICAL SUPPORT

For troubleshooting, information or assistance, please go online to [www.cohesionbio.com](http://www.cohesionbio.com) or contact us at [techsupport@cohesionbio.com](mailto:techsupport@cohesionbio.com)

## IX. NOTES